

【文献综述】

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1.1 EGCG 的生物学特性

茶多酚 (tea polyphenols, TP) 是茶叶中多酚类物质的总称, 包括黄烷醇类、花色苷类、黄酮类、黄酮醇类和酚酸类等, 其中以黄烷醇类物质 (儿茶素) 最为重要, 占 TP 总量的 65% ~ 80%, 主要含有 4 种单体: EGCG、表儿茶素没食子酸酯 (epieabechingallate, ECG)、表儿茶素 (epicatechin, EC) 及表没食子儿茶素 (epigallocatechin, EGC)。其中, EGCG 含量最高, 占儿茶素的 80% 左右, 也是茶多酚中生物活性最强的成分^[1]。研究表明, EGCG 能够保护细胞和 DNA 免受损害, 具有显著的抗氧化、清除体内自由基、抗突变、抗病毒、调节机体免疫功能、诱导肿瘤细胞凋亡及光保护等一系列生物学活性^[2-3]。由于 EGCG 在绿茶中的含量较高, 同时具有非常强的抗氧化活性, 近年来, EGCG 得到广泛的研究, 并且成为一种受人们欢迎的药品。研究

表明,EGCG可以预防慢性疾病,如神经变性疾病、肥胖、癌症、2型糖尿病、动脉粥样硬化及心脏疾病等^[4-8]。同时,EGCG能够保护心脏、肾脏、神经系统、角膜、视网膜及晶状体等,减轻氧化对它们的损伤^[9-14]。

1.2 EGCG抗氧化和抗凋亡作用 研究表明,EGCG具有抗氧化和抗凋亡的作用^[15]。EGCG分子中含有8个酚性羟基结构,具有淬灭单线态氧、清除自由基的能力,其抗氧化活性是超氧化物歧化酶(superoxide dismutase, SOD)的6倍,是维生素E的20倍^[16]。它还可能通过清除活性氧(reactive oxygen species, ROS)^[17],抑制细胞内生物大分子的过氧化,进而改变细胞内氧化还原状态,从而抑制细胞凋亡。EGCG发挥抗氧化作用的机制主要包括:(1)清除自由基;(2)增加抗氧化酶的活性;(3)络合诱导氧化的金属离子;(4)促进体内抗氧化物再生。Park等^[18]研究发现,EGCG可以通过清除ROS,调节Bcl-2家族基因及caspases的表达、抑制NO诱导的牙釉质细胞的凋亡;Zhou等^[19]研究发现,EGCG可以通过清除ROS及减少线粒体损伤来抑制血管紧张素II诱导的人脐静脉血管内皮细胞的凋亡;Saito等^[20]认为EGCG可以通过抑制氧化应激引起的DNA损伤及通过上调血红素氧合酶-1和Bcl-2的表达来抑制细胞凋亡。Adikesavan等^[21]研究发现,EGCG可以减少氧化应激对心肌的损伤,同时可以通过抑制线粒体介导的细胞凋亡保护心脏。

1.3 EGCG对紫外线诱导损伤的保护作用 紫外线辐射性氧化损伤的病理机制可概括为:紫外线照射产生的ROS如单线氧、超氧阴离子和羟自由基等,可破坏DNA结构,激活蛋白激酶。ROS首先损伤细胞,诱发细胞DNA发生改变、蛋白质和脂质损伤、改变细胞膜通透性,影响酶的活性,最终导致细胞凋亡。

细胞内ROS的平衡依赖于细胞内正常新陈代谢和抗氧化系统清除能力之间的动态平衡。机体内的抗氧化系统包括:非酶系统[如谷胱甘肽(glutathione, GSH)、胆红素、维生素E和维生素C]和酶促反应系统[如SOD、过氧化氢酶、谷胱甘肽过氧化物酶(GSH-Px)以及谷胱甘肽还原酶(GSH-Pb)]。机体内抗氧化系统与氧化物之间的失衡,将对机体造成损伤。

EGCG对紫外线诱导的细胞损伤具有保护作用。EGCG可通过调节micro-RNA的表达来抵抗紫外线B对人皮肤成纤维细胞的损伤^[22]。EGCG还可以通过清除活性氧、减少黑色素生成,减轻紫外线对HaCaT细胞的损伤^[23]。Chaudhury等^[24]研究证实EGCG可以通过抗氧化应激作用保护人 γ B-晶状体蛋白免受紫外线的损伤。

2 EGCG抗氧化与光保护作用在眼科的应用

由于EGCG的抗氧化作用和光保护作用,近年

来一些学者专注于EGCG对眼科疾病防治的研究。

2.1 EGCG对视网膜色素上皮细胞的保护作用

Cao等^[25]发现EGCG可以通过抑制氧化应激及调节JNK1/c-Jun通路来保护紫外线B诱导的ARPE19细胞的凋亡。EGCG通过升高survivin基因的表达,减轻线粒体功能紊乱和减少DNA碎片,从而保护紫外线B对视网膜色素上皮细胞的损伤^[26]。EGCG可以调节视网膜色素上皮细胞的自我吞噬作用来减少紫外线B对视网膜的损伤^[27]。有研究表明,EGCG可以减轻H₂O₂诱导的鼠视网膜色素上皮的损伤^[28],这表明,EGCG在预防与H₂O₂诱导氧化损伤相关的视网膜疾病方面有潜在作用。

2.2 EGCG对神经节细胞的保护作用 EGCG具有明显的神经保护作用,其作用机制与EGCG的抗氧化作用有关^[29-30]。目前关于EGCG对视神经的保护作用主要集中在其对视网膜神经节细胞(retinal ganglion cells, RGC)的保护作用。有研究表明,一定浓度的EGCG对H₂O₂、紫外线照射诱导的RGC-5的氧化损伤有保护作用^[31-32]。姜利斌等^[33]研究发现,EGCG对大鼠视神经钳伤后RGC具有一定的保护作用,并推测EGCG可能是通过抗氧化、清除自由基等作用而保护视神经。

2.3 EGCG对晶状体上皮细胞的保护作用 有学者发现EGCG可抑制地塞米松损伤所诱导的兔晶状体上皮细胞凋亡,并推测EGCG可以通过清除自由基使体内抗氧化体系再生,抑制过氧化反应,从而抑制细胞凋亡^[34]。同时,一些研究发现EGCG对紫外线B造成的晶状体上皮细胞的损伤有保护作用^[28],EGCG可以减少紫外线B诱导的人晶状体上皮细胞内ROS的产生及NADPH氧化酶的活性^[35-36],对DNA的损伤有拮抗作用^[37],为EGCG用于白内障的防治提供新的思路。司南等^[38]应用紫外线B照射新西兰大白兔后给予不同浓度的EGCG滴眼液,发现EGCG滴眼液可以进入兔眼房水发挥抗氧化作用,其中以浓度为200 $\mu\text{mol} \cdot \text{L}^{-1}$ 的EGCG滴眼液的抗氧化作用效果最佳。EGCG可以通过抑制caspase-3、caspase-9的表达^[39]或抑制线粒体通路介导的细胞凋亡保护晶状体上皮细胞免受过氧化氢的损伤^[40]。EGCG可能通过提高细胞内SOD、GSH-Px含量,降低MDA含量,发挥其较强的抗氧化作用^[14];进而通过调节Bcl-2/Bax、p53、c-fos和c-myc的表达^[41],抑制高糖所诱导的晶状体上皮细胞的凋亡。

2.4 EGCG对角膜的保护作用 角膜碱烧伤后白细胞激活、炎症细胞浸润和释放大量氧自由基是导致角膜损伤的主要途径,炎症反应与氧化反应又相互影响,且炎症也与新生血管有密切关系;EGCG可有效促进碱烧伤后鼠角膜上皮修复,抑制新生血管形成和炎症细胞浸润^[42-43]。联合抗真菌和抗氧化治疗可减少真菌性角膜炎的氧化应激。有研究表明,

联合应用伏立康唑和 EGCG,与单用伏立康唑相比,前者可明显减轻真菌性角膜炎所引起的炎症反应^[44]。EGCG 可有效抑制人角膜缘上皮细胞内 ROS 的产生,从而保护角膜缘上皮细胞免受氧化损伤^[45],为 EGCG 在干眼的防治提供了依据。此外,EGCG 可增加抗氧化系统酶的活性及抑制脂质过氧化反应和蛋白质氧化来保护紫外线 B 对角膜的损伤^[46]。

3 结论

EGCG 具有多种生物活性,其抗氧化作用一直是人们研究的重点。近年来一些学者专注于 EGCG 对眼科疾病防治的研究。一定浓度的 EGCG 对视网膜上皮细胞、视网膜神经节细胞、晶状体上皮细胞及角膜细胞等均有保护作用,为 EGCG 在眼科的应用提供了依据。

参考文献

[1] HARATIFAR S, CORREDIG M. Interactions between tea catechins and casein micelles and their impact on renneting functionality[J]. *Food Chem*, 2014, 143(1): 27-32.

[2] AN Z, QI Y, HUANG D, GU X, TIAN Y, LI P, et al. EGCG inhibits Cd²⁺-induced apoptosis through scavenging ROS rather than chelating Cd²⁺ in HL-7702 cells[J]. *Toxicol Mech Methods*, 2014, 24(4): 259-267.

[3] CHUNG S S, VADGAMA J V. Curcumin and epigallocatechin gallate inhibit the cancer stem cell phenotype via down-regulation of STAT3-NFκB signaling[J]. *Anticancer Res*, 2015, 35(1): 39-46.

[4] BITU PINTO N, DA SILVA ALEXANDRE B, NEVES KR, SILVA AH, LEAL LK, VIANA GS. Neuroprotective properties of the standardized extract from camellia sinensis (green tea) and its main bioactive components, epicatechin and epigallocatechin gallate, in the 6-OHDA model of parkinson's Disease[J]. *Evid-bas Com Alt Med*, 2015, 2015: 161092.

[5] LI X, LI S, CHEN M, WANG J, XIE B, SUN Z. (-)-Epigallocatechin-3-gallate (EGCG) inhibits starch digestion and improves glucose homeostasis through direct or indirect activation of PXR/CAR-mediated phase II metabolism in diabetic mice[J]. *Food Funct*, 2018, 9(9): 4651-4663.

[6] YANG C S, ZHANG J, ZHANG L, HUANG J, WANG Y. Mechanisms of body weight reduction and metabolic syndrome alleviation by tea[J]. *Mol Nutr Food Res*, 2016, 60(1): 160-174.

[7] YU C, JIAO Y, XUE J, ZHANG Q, YANG H, XING L, et al. Metformin sensitizes non-small cell lung cancer cells to an epigallocatechin-3-gallate (EGCG) treatment by suppressing the Nrf2/HO-1 signaling pathway[J]. *Int J Biol Sci*, 2017, 13(12): 1560-1569.

[8] ENG Q Y, THANIKACHALAM P V, RAMAMURTHY S. Molecular understanding of epigallocatechin gallate (EGCG) in cardiovascular and metabolic diseases[J]. *J Ethnopharmacol*, 2018, 210: 296-310.

[9] ARAS-LÓPEZ R, ALMEIDA L, ANDREU-FERNÁNDEZ V, TOVAR J, MARTÍNEZ L. Anti-oxidants correct disturbance of redox enzymes in the hearts of rat fetuses with congenital diaphragmatic hernia[J]. *Pediatr Surg Int*, 2018, 34(3): 307-313.

[10] GAO Z, HAN Y, HU Y, WU X, WANG Y, ZHANG X, et al. Targeting HO-1 by Epigallocatechin-3-Gallate reduces contrast-induced renal injury via anti-oxidative stress and anti-inflammation pathways[J]. *PLoS One*, 2016, 11(2): e0149032.

[11] KIAN K, KHALATBARY AR, AHMADVAND H, KARIMPOUR MALEKSHAH A, SHAMS Z. Neuroprotective effects of (-)-epigallocatechin-3-gallate (EGCG) against peripheral nerve transection-induced apoptosis[J]. *Nutr Neurosci*, 2018, 2: 1-9.

[12] CAVET M E, HARRINGTON K L, VOLLMER T R, WARD K W, ZHANG J Z. Anti-inflammatory and anti-oxidative effects

of the green tea polyphenol epigallocatechin gallate in human corneal epithelial cells[J]. *Mol Vis*, 2011, 17: 533-542.

[13] YANG Y, QIN Y J, YIP Y W, CHAN K P, CHU K O, CHU W K, et al. Green tea catechins are potent anti-oxidants that ameliorate sodium iodate-induced retinal degeneration in rats[J]. *Sci Rep*, 2016, 6: 29546.

[14] CHEN T, LIU P, WANG J X, SHAN D, CHE W, ZHANG L J. Effects of EGCG on hyperglycemic-induced oxidative stress in human lens epithelial cells[J]. *Int J Ophthalmol*, 2016, 16(6): 1029-1031.

[15] FILIP A, DAICOVICIU D, CLICHICI S, MOCAN T, MURESAN A, POSTESCU I D. Photoprotective effects of two natural products on ultraviolet B-induced oxidative stress and apoptosis in SKH-1 mouse skin[J]. *J Med Food*, 2011, 14(7-8): 761-766.

[16] PABARI A, YANG S Y, MOSAHEBI A, SEIFALIAN A M. Recent advances in artificial nerve conduit design: strategies for the delivery of luminal fillers[J]. *J Control Release*, 2011, 156(1): 2-10.

[17] ZHANG W, LV J, ZHANG Y, JIANG Y, CHU C, WANG S. Epigallocatechin gallate promotes the development of mouse 2-cell embryos *in vitro* by regulating mitochondrial activity and expression of genes related to p53 signalling pathway[J]. *Basic Clin Pharmacol Toxicol*, 2014, 115(5): 403-410.

[18] PARK S Y, JEONG Y J, KIM S H, JUNG J Y, KIM W J. Epigallocatechin gallate protects against nitric oxide-induced apoptosis via scavenging ROS and modulating the Bcl-2 family in human dental pulp cells[J]. *J Toxicol Sci*, 2013, 38(3): 371-378.

[19] ZHOU X, LIANG L, ZHAO Y, ZHANG H. Epigallocatechin-3-Gallate ameliorates angiotensin II-induced oxidative stress and apoptosis in human umbilical vein endothelial cells through the activation of nrf2/caspase-3 signaling[J]. *J Vasc Res*, 2017, 54(5): 299-308.

[20] SAITO K, MORI S, DATE F, ONO M. Epigallocatechin gallate inhibits oxidative stress-induced DNA damage and apoptosis in MRL-Fas (lpr) mice with autoimmune sialadenitis via up-regulation of heme oxygenase-1 and Bcl-2[J]. *Autoimmunity*, 2014, 47(1): 13-22.

[21] ADIKESAVAN G, VINAYAGAM M M, ABDULRAHMAN L A, CHINNASAMY T. (-)-Epigallocatechin-gallate (EGCG) stabilize the mitochondrial enzymes and inhibits the apoptosis in cigarette smoke-induced myocardial dysfunction in rats[J]. *Mol Biol Rep*, 2013, 40(12): 6533-6545.

[22] AN I S, AN S, PARK S, LEE S N, BAE S. Involvement of microRNAs in epigallocatechin gallate-mediated UVB protection in human dermal fibroblasts[J]. *Oncol Rep*, 2013, 29(1): 253-259.

[23] KIM E, HWANG K, LEE J, HAN S Y, KIM E M, PARK J, et al. Skin protective effect of epigallocatechin gallate[J]. *Int J Mol Sci*, 2018, 19(1): 173.

[24] CHAUDHURY S, BAG S, BOSE M, DAS A K, GHOSH A K, DASGUPTA S. Protection of human γB-crystallin from UV-induced damage by epigallocatechin gallate: spectroscopic and docking studies[J]. *Mol Biosyst*, 2016, 12(9): 2901-2909.

[25] CAO G, CHEN M, SONG Q, LIU Y, XIE L, HAN Y, et al. EGCG protects against UVB-induced apoptosis via oxidative stress and the JNK1/c-Jun pathway in ARPE19 cells[J]. *Mol Med Rep*, 2012, 5(1): 54-59.

[26] XU J Y, WU L Y, ZHENG X Q, LU J L, WU M Y, LIANG Y R. Green tea polyphenols attenuating ultraviolet B-induced damage to human retinal pigment epithelial cells *in vitro*[J]. *Invest Ophthalmol Vis Sci*, 2010, 51(12): 6665-6670.

[27] LI C P, YAO J, TAO Z F, LI XM, JIANG Q, YAN B. Epigallocatechin-gallate (EGCG) regulates autophagy in human retinal pigment epithelial cells: a potential role for reducing UVB light-induced retinal damage[J]. *Biochem Biophys Res Commun*, 2013, 438(4): 739-745.

[28] CIA D, VERGNAUD-GAUDUCHON J, JACQUEMOT N, DOLY M. Epigallocatechin gallate (EGCG) prevents H₂O₂-induced oxidative stress in primary rat retinal pigment epithelial cells[J]. *Curr Eye Res*, 2014, 39(9): 944-952.

[29] SHEN C, CHEN L, JIANG L, LAI T Y. Neuroprotective effect of epigallocatechin-3-gallate in a mouse model of chronic glaucoma[J]. *Neurosci Lett*, 2015, 600: 132-136.

[30] FAN B, LI G Y, LI Y P, CUI J Z. Neuroprotective effect of epigallocatechin gallate on oxidative-stress-injured retinal cells[J]. *Zhonghua Yi Xue Za Zhi*, 2008, 88(24): 1711-

- 1714.
- [31] ZHENG B, LI G Y, LI J R, ZHENG Y C. Oxidative and antioxidant effects of green tea extract EGCG on visual ganglion cells rgc-5[J]. *Chin J Ger Med*, 2014, 34: 3924-3926.
- 郑斌, 李光宇, 李佳睿, 郑永晨. 绿茶提取物 EGCG 对视神经节细胞 RGC-5 的促氧化和抗氧化作用[J]. 中国老年杂志, 2014, 34: 3924-3926.
- [32] JIN J, YING H, HUANG M, DU Q. Bioactive compounds in green tea leaves attenuate the injury of retinal ganglion RGC-5 cells induced by H₂O₂ and ultraviolet radiation[J]. *Pak J Pharm Sci*, 2015, 28(6 Suppl): 2267-2272.
- [33] JIANG L B, XIE J, ZHANG T, JIN Y L, YANG D M, CHEN F. Protective effect of epigallocatechin gallate on retinal ganglion cells after optic nerve forceps injury in rats[J]. *Chin J Fund Dis*, 2011, 27(1): 74-78.
- 姜利斌, 谢君, 张婷, 金玉兰, 杨冬梅, 陈菲. 表没食子儿茶素没食子酸酯对大鼠视神经钳夹伤后视网膜神经节细胞的保护作用[J]. 中华眼底病杂志, 2011, 27(1): 74-78.
- [34] LIU X P, HE X Z. Effects of epigallocatechin gallate on inhibition of dexamethasone induced cataract *in vitro* in rabbits[J]. *Int J Ophthalmol*, 2010, 10(3): 443-445.
- [35] YAO J, LIU Y, WANG X, SHEN Y, YUAN S, WAN Y, et al. UVB radiation induces human lens epithelial cell migration via NADPH oxidase-mediated generation of reactive oxygen species and up-regulation of matrix metalloproteinases[J]. *Int J Mol Med*, 2009, 24(2): 153-159.
- [36] LIU Y, YANG W, CHEN M R, YAO J, JIANG Q. Experimental study on inhibition of uvb-induced lens epithelial cell damage by epigallocatechin gallate[J]. *Rec Adv Ophthalmol*, 2011, 31(6): 516-518, 523.
- 刘媛, 杨文, 陈美蓉, 姚进, 蒋沁. 表没食子儿茶素没食子酸酯抑制 UVB 介导的晶状体上皮细胞损伤的实验研究[J]. 眼科新进展, 2011, 31(6): 516-518, 523.
- [37] WU Z H, WANG M R, YAN Q C, PU W, ZHANG J S. Analysis of uv-induced DNA damage in human lens epithelial cells and the antagonistic effects of antioxidants on DNA damage[J]. *Chin J Ophthalmol*, 2006, 42(11): 1002-1007.
- 吴志鸿, 王冕蓉, 阎启昌, 濮伟, 张劲松. 羟星法分析紫外线诱导的人晶状体上皮细胞 DNA 的损伤及抗氧化剂对 DNA 损伤的拮抗作用[J]. 中华眼科杂志, 2006, 42(11): 1002-1007.
- [38] SI N, WU Z H, MA Y, ZHOU L D, ZHENG J C. Study on the antioxidant effect of epigallocatechin gallate eye drops in rabbit eyes[J]. *Chin J Emerg Res Dis Med*, 2016, 4(11): 386-389.
- 司南, 吴志鸿, 马艳, 周立冬, 郑静晨. 表没食子儿茶素没食子酸酯滴眼液在兔眼中抗氧化作用的研究[J]. 中国急救复苏与灾害医学杂志, 2016, 4(11): 386-389.
- [39] YANG Z, LIU P, WANG J X, SHAN D, CHE W, ZHANG L J. Protective effect of EGCG on H₂O₂ induced oxidative damage of HLE cells[J]. *Adv Mod Biom Sci*, 2016, 28: 5410-5413, 5427.
- 杨哲, 刘平, 王嘉翔, 单多, 车雯, 张丽娟. EGCG 对 H₂O₂ 诱导 HLE 细胞氧化损伤的保护作用[J]. 现代生物医学进展, 2016, 28: 5410-5413, 5427.
- [40] YAO K, YE P, ZHANG L, TAN J, TANG X, ZHANG Y. Epigallocatechin gallate protects against oxidative stress-induced mitochondria-dependent apoptosis in human lens epithelial cells[J]. *Mol Vis*, 2008, 14: 217-223.
- [41] YE P, LIN K, LI Z, LIU J, YAO K, XU W. (-)-Epigallocatechin gallate regulates expression of apoptotic genes and protects cultured human lens epithelial cells under hyperglycemia[J]. *Mol Biol (Mosk)*, 2013, 47(2): 251-257.
- [42] ZHANG W L, WU Z H. Study on inhibition of corneal neovascularization by EGCG liposomes[J]. *Chin J Emerg Res Dis Med*, 2017, 12(1): 31-36.
- 张伟莉, 吴志鸿. EGCG 脂质体抑制大鼠角膜新生血管的研究[J]. 中国急救复苏与灾害医学杂志, 2017, 12(1): 31-36.
- [43] WU L Q, LU M. Therapeutic effect of epigallocatechin gallate on corneal alkali burn in mice[J]. *J Zhejiang Univ (Med Ed)*, 2015, 44(1): 15-23.
- 吴联群, 卢敏. 表没食子儿茶素没食子酸酯对小鼠角膜碱烧伤的治疗作用[J]. 浙江大学学报(医学版), 2015, 44(1): 15-23.
- [44] RUBAN V V, ARCHANA P T, SUNDARARAJAN M, GERALDINE P, THOMAS P A. Inflammation and oxidative stress in corneal tissue in experimental keratitis due to *Fusarium solani*: Amelioration following topical therapy with voriconazole and epigallocatechin gallate[J]. *Mycoses*, 2018, 61(3): 159-171.
- [45] STODDARD A R, KOETJE L R, MITCHELL A K, SCHOTANUS M P, UBELS J L. Bioavailability of antioxidants applied to stratified human corneal epithelial cells[J]. *J Ocul Pharmacol Ther*, 2013, 29(7): 681-687.
- [46] MU-HSIN CHEN, CHIA-FANG TSAI, YU-WEN HSU, FUNG-JOU LU. Epigallocatechin gallate eye drops protect against ultraviolet B-induced corneal oxidative damage in mice[J]. *Mol Vis*, 2014, 20: 153-162.
- [12] EUSTIS H S, ROWLAND A P. Corneal abrasions associated with Crawford mono-canalicular tubes[J]. *J Pediatr Ophthalmol Strab*, 2011, 48: e61-62.
- [13] ZHANG J X, DENG H W, YE L. Technology of lacrimal intubation[J]. *Chin J Ophthalmol*, 2011, 47(8): 765-767.
- 张敬先, 邓宏伟, 叶琳. 泪道置管技术[J]. 中华眼科杂志, 2011, 47(8): 765-767.
- [14] XU J, HONG J, SUN X, LIU Z, MASHAGHI A, INOMATA T, et al. Combined lacrimal passage probing and tobramycin/dexamethasone ophthalmic ointment infiltration: a minimally invasive surgical procedure for incomplete nasolacrimal duct obstruction[J]. *Medicine*, 2015, 94(36): e1483.
- [15] YU G, HU M, WU Q, CAO W H, FAN Y W, LIN Q, et al. Factors affected therapeutic results in treatment of children congenital nasolacrimal duct obstruction by Ritleng lacrimal intubation[J]. *Chin J Ophthalmol*, 2012, 48(5): 423-427.
- 于刚, 胡曼, 吴倩, 曹文红, 樊云葳, 蔺琪, 等. Ritleng 泪道插管术治疗儿童先天性鼻泪管阻塞疗效的影响因素分析[J]. 中华眼科杂志, 2012, 48(5): 423-427.
- [16] MADER T H, WELLS J R, ROCKWELL J C. A method of removing displaced silicone tubing from the nasolacrimal duct system[J]. *Am J Ophthalmol*, 1985, 99(6): 730-731.
- [17] LI F M, XIE L X. Chinese ophthalmology[M]. Beijing: People's Medical Publishing House, 2014: 154-155.
- 李凤鸣, 谢立信. 中华眼科学[M]. 北京: 人民卫生出版社, 2014: 154-155.
- [18] MERBS S L, HARRIS L L, IWAMOTO M A, ILIFF N T. Prevention of prolapsed silicone stents in lacrimal intubation using an intrasac fixation suture[J]. *Arch Ophthalmol*, 1999, 117(8): 1092-1095.
- [19] HAKIM O M, EL-HAG Y G. Silicone intubation with trans-sac fixation to prevent tube dislocation[J]. *J AAPOS*, 2005, 9(6): 558.

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- [3] LIN L, YANG L, JIN X, ZHAO Y, FAN F. Management lacrimal sac abscesses using lacrimal probe and crawford silicon tube[J]. *BMC Ophthalmol*, 2016, 16(1): 211.
- [4] WANG L, CHEN D, WANG Z. New technique for lacrimal system intubation[J]. *Am J Ophthalmol*, 2006, 142(2): 252-258.
- [5] DEMIRCI H, ELNER V M. Double silicone tube intubation for the management of partial lacrimal system obstruction[J]. *Ophthalmology*, 2008, 115(2): 383-385.
- [6] LIANG X, LIN Y, WANG Z, LIN L, ZENG S, LIU Z, et al. A modified bicanalicular intubation procedure to repair canalicular lacerations using silicone tubes[J]. *Eye*, 2012, 26(12): 1542-1547.
- [7] MIMURA M, UEKI M, OKU H, SATO B, IKEDA T. Indications for and effects of Nunchaku-style silicone tube intubation for primary acquired lacrimal drainage obstruction[J]. *Jpn J Ophthalmol*, 2015, 59(4): 1-7.
- [8] CHOI S C, CHOI H S, JANG J W, KIM S J, LEE J H. Comparison of the efficacies of 0.94 mm and double silicone tubes for treatment of canalicular obstruction[J]. *Korean J Ophthalmol*, 2017, 31(1): 1-8.
- [9] CHA D S, LEE H, PARK M S, LEE J M, BAEK S H. Clinical outcomes of initial and repeated nasolacrimal duct office-based probing for congenital nasolacrimal duct obstruction[J]. *Korean J Ophthalmol*, 2010, 24(5): 261-266.
- [10] CAKMAK S S. A new method for removing displaced silicone tubing from the nasolacrimal duct[J]. *Orbit*, 2009, 28(6): 444-446.
- [11] MIMURA M, UEKI M, OKU H, SATO B, IKEDA T. Evaluation of granulation tissue formation in lacrimal duct post silicone intubation and its successful management by injection of prednisolone acetate ointment into the lacrimal duct[J]. *Jpn J Ophthalmol*, 2016, 60(4): 280-285.